



Asia Hepato Gene Co. Product Datasheet

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pHX TA Cloning kit

10 Reactions

Application:

1. PCR product cloning
2. Multiple cloning region

Kit Components:

	Item	Concentration	Amount	Storage
1	pHX TA vector	<ul style="list-style-type: none"> · Size: 3,000bp · 50ng/μl · Ampicillin resistance · Multiple cloning region 	25μl	-20°C
2	10x Ligation Buffer A	0.4M Tris-HCl, 0.1mM MgCl ₂ , 0.1M DTT, 5mM ATP	10μl	-20°C
3	10x Ligation Buffer B	Enhance ligation efficiency	10μl	-20°C
4	Ligase	50mM KCl, 20mM Tris-HCl.(pH7.5).0.1mM EDTA, 1mM DTT, 50% glycerol	5μl	-20°C
5	SOC Medium	2% Tryptone, 0.5% Yeast Extract, 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl ₂ , 10 mM MgSO ₄ , 20 mM glucose.	1.5ml	25°C
6	T7 Forward primer	10μM/μl in 1×TE buffer, pH8.0 ,Tm 50°C Sequence: 5'-TAATACGACTCACTATAGGG-3'	50μl	-20°C
7	T7 Reverse primer	10μM/μl in 1×TE buffer, pH8.0, Tm 50°C Sequence: 5'-ATCCGGATATAGTTCCTCCTTTC-3'	50μl	-20°C
8	DH5α competent cell	Transformation efficiency >3x10 ⁸ cfu/μg DNA	100μl×10	-80°C
9	pUC19 Control DNA	10 ⁻⁴ μg/μl	5μl	-20°C

PCR Primer Design:

Forward primer	-
Reverse primer	-

High Yield PCR Product:

94°C → 94°C → X°C → 72°C → 72°C → 25°C
 5min 30sec 30sec 30sec 7min end
30 cycles

Load 16μl PCR product to run 1-2% small wells agarose gel, target gene should be clearly visible on the gel

For research use only, not for diagnostic or therapeutic use.

Ligation:

1. Add as following and carefully flick the tube 5-6 times to mix. Do not vortex.
* PCR product contain Ampicillin resistance template must be purified by agarose gel to remove Ampicillin resistance template

pHX TA vector	2.5µl	2.5µl	2.5µl	2.5µl
PCR product <3kb	1µl	2µl	3µl	4µl
10x Ligation Buffer A	1µl	1µl	1µl	1µl
10x Ligation Buffer B	1µl	1µl	1µl	1µl
Ligase	0.5µl	0.5µl	0.5µl	0.5µl
dH2O	4µl	3µl	2µl	1µl
total	10µl	10µl	10µl	10µl

2. The optimal ligation efficiency molar ratio vector : insert =1 : 3.
Low yield and long PCR product need more.
3. Centrifuge briefly and incubate at room temperature (25°C) for 5-20 minutes will suffice for PCR product <3kb.
4. If PCR product >3kb, incubate at 4°C overnight

Transformation:

1. Thaw DH5α competent cell at room temperature before use
2. Add 108µl competent cell into 10µl ligation mixture and mix gently.
3. On ice or 4°C for 10 minutes
4. 42°C 45 seconds
5. Add 80µl SOC Medium
6. Spread 168µl cells onto 20-50µg/ml Ampicillin plate. Excessive antibiotic significantly decreased the transformation efficiency
7. 37°C 12-16 hours

Analyzing Positive Clones:

1. Pick 12 colonies or more and culture them overnight in LB containing 20-50 µg/ml ampicillin.
2. Isolate plasmid DNA.
3. Analyze the plasmids by restriction analysis, sequencing,
4. PCR use **target gene forward primer** and **T7 Reverse primer** (add 0.5µl per 25µl PCR reaction) to confirm the presence and correct orientation of the insert.

Long-Term Storage:

Keep the correct clone in 20-30% glycerol and store at -80°C.

Purification of Restriction Enzyme Cut Fragment

Extract plasmid from 1ml LB overnight culture. After enzyme cutting plasmid & RNAsA digest RNA, running 1-2% agarose gel and cutting target DNA fragment from the gel.

Purify the DNA fragment in the gel by PCR Product Purification Kit.

PHX TA vector cloning site:

T7 forward primer

1 TAATACGACT CACTATAGGG A GACCACAA C GTTTCCCTCT AGAA AT AA TTTTGTTTAA C TTTAAGAAG
70 GAGATATACA T ATG CGG GGT TCT CAT CAT CAT CAT CAT GGT ATG GCT AGC ATG ACT GGT GG
134 AC AGC AAA TG G GTC GGG A TC TGT ACG A CG ATG ACG ATA AGG A TCGAT GGG|GA TC CGA G C|TCG
196 AGCGGCCGC C AGTGTGATGG ATATCTGCA|G|AATT CGC CCT TTC|CAT GGT T PCR product A CACTTGGT
254 AAGGGCG|AA TT CCAGCACACT GGCGGCCGTT ACTAGTG|GAT CCGAGC T|CGG TAC|CA|AG CTT GATC
319 CGGCTG CTA ACAAAGC CCGAA AGGA AGCTGA GTTG GCTGCGCCAC CG CTGAGCAA TAACT AGCAT

T7 reverse priming site

The phenotypes of protein toxicity in molecular biology studies

- Toxic protein cloning problems
 - No colony in cloning
 - Fewer colonies than those in regular cloning experiments
 - Small percentage of positive clones
 - Wrong orientation
 - Mutations leading to defective products or no expression
- Toxic protein expression problems
 - No expression
 - Low yield
 - Defective proteins
 - The expression seems inconsistent or unpredictable.
- Toxic protein host cell growth rate and cell density problems
 - Cell grows significantly slower to reach its normal density before or after induction.
- Transformation efficiency problems
 - The transformation efficiency of the vector containing a toxic protein is lower than the control